Atropisomerism Observed in Indometacin Derivatives

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ABSTRACT



To elucidate the active conformation of indometacin that differentiates between cyclooxygenase-1 (COX-1) and cyclooxygenase-2 (COX-2), the stereochemistry around the *N*-benzoylated indole moiety of indometacin was studied. Resolution of stable atropisomers as representative conformations was found to be possible by restricting rotation about the N–C7' and/or C7'–C1' bond. Only the a*R*-isomer showed specific inhibition of COX-1, and COX-2 was not inhibited by either atropisomer.

Indometacin, a "classical" nonsteroidal antiinflammatory drug (NSAID), acts at the cyclooxygenase (COX) active site and inhibits two isoforms of COX, i.e., COX-1 and COX-2, with little specificity, leading to serious side effects.^{1–7} Therefore COX-1/2-selective indometacin

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analogues have been extensively explored in attempts to develop a new generation of NSAIDs.⁸⁻¹¹

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In the design of new COX-1/2-selective indometacin analogues, the conformation of the *N*-benzoyl moiety has been considered. The benzoyl (phenyl) group can adopt either a *cis* or *trans* conformation with respect to the indole (Figure 1a). In 1996, Loll et al. proposed that indometacin interacts with COX-1 in two possible binding modes corresponding to the *cis* or *trans* form.¹² Independently, the group of Miyashiro and Penning reported that indometacin adopts a *cis* conformation in its complex with COX-2.¹³ Since then, the *cis/trans* conformations of

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Figure 1. (a) Conformations traditionally used for indometacin. (b) 2',6'-Disubstituted indometacin derivative **A**.

indometacin have been regarded as a key feature for improved drug design.^{14,15} However, "*cis/trans*" terminology is associated with a planar structure in which the indole and benzoyl (phenyl) groups are coplanar. Such vague terminology could prove misleading in the present case. X-ray crystallographic analysis has shown that indometacin, even in a crystal state, adopts several different conformations, in which the indole and benzoyl (phenyl) groups are twisted relative to each other.^{16,17}

Similarly, in solution the 1-benzoyl-2-methylindole moiety of indometacin is not expected to adopt a completely planar conformation because of the large steric hindrance between the phenyl and indolyl groups. The N-benzoyl moiety has two sp^2-sp^2 axes along the N-C7' and C7'-C1' bonds that can provide numerous conformations of indometacin in solution. To elucidate the active conformation of indometacin, the resolution of stable atropisomers as a representative conformation by restricting the rotation of the N-C7' and/or C7'-C1' bond could provide a new means to study the molecular origins of COX-1/2selectivity. At present, atropisomerism is most common in biphenyl or biaryl C-C bond stereochemistry, although atropisomers arising from C–N bonds have recently attracted much attention.¹⁸⁻²⁵ However, to the best of our knowledge, reports on the chirality of N-benzovlindoles, with chirality arising from the two sp^2-sp^2 axes of the aromatic N-CO-Ar bonds, have not yet been

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published. Here we show presence of atropisomers based on the C7'-C1' axis in the *N*-benzoyl moiety of 2',6'disubstituted indometacin derivatives (**A**, Figure 1b), which possess high stereochemical stability and are recognized by COX-1. This novel chirality is produced by fixing the C7'-C1' axis alone, and hence the other N-C7' axis independently rotates to form an equilibrium between *cis*and *trans*-like conformations in each enantiomer.





Through the examination of various indometacin derivatives, we found that the C-2' and C-6' disubstituted indometacin derivatives (1-4, Scheme 1) exist in *cis-trans* equilibrium: the ¹H NMR spectrum of 1 in CDCl₃ was observed as two sets of resonances in a 1:1.7 ratio (see Supporting Information). In order to assign the *cis/trans* conformations, the NOESY method was used for the trisubstituted (at C-2', C-4', and C-6') benzoyl derivative 2. In the NOESY specrum of the minor form, H7 showed a correlation peak to H3'/5', which confirmed the position of the benzoyl (phenyl) ring to be *cis*. On the contrary, for the major isomer of 2, 2-CH₃ showed a correlation peak to H3'/5', which confirmed the position of the benzoyl ring to be trans (see Supporting Information). We therefore reasoned that compound 1 exists as a mixture of cis-like and *trans*-like²⁶ forms (*cis/trans* = 1:1.7) in solution. Compound 1 was observed as an inseparable single peak on nonchiral HPLC at room temperature, indicating that conformers are at equilibrium as the result of rotation around the N–C7' axis.²⁷ For 1-4, either or both conformations (cis-like and trans-like) could be adopted when it binds to an enzyme. However, the ratio of cis-like and *trans*-like (1:1.6-1.7) may be taken to indicate a tendency to adopt the trans-like conformer.

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⁽²⁶⁾ These "*trans*-like/*cis*-like" terms have the implication that the carbonyl group takes a type of perpendicular position with respect to the plane of the phenyl group of the benzoyl moiety.

⁽²⁷⁾ In order to examine the equilibrium between the *cis-/trans*-conformers, VT-NMR has been measured (see: Supporting Information).



Figure 2. Structure of 3 determined using X-ray crystallographic analysis.

After confirming that the N-C7' axis of N-benzoylated indole derivatives allows rotation in solution, the next step was to examine the C7'-C1' axis. The structural analysis was supported by single-crystal X-ray analysis of 3, in which the dihedral angle (ϕ) C2'-C1'-C7'-O was shown to be 67.8°, confirming that the benzene ring is essentially orthogonal to the indole ring (Figure 2). This geometry convinced us that the atropisomeric property arises from the C7'-C1' axis in C-2' and C-6' disubstituted indole derivatives. As expected, compound 4, in which Cl and I are present at C-2' and C-6', respectively, gave important insight into this. On the basis of the ¹H NMR spectrum, it is clear that **4** also exists as an equilibrium mixture of *cis*-like and *trans*-like conformation (*cis*-like:*trans*-like = 1:1.6) in $CDCl_3$. It was also observed as a single peak on nonchiral HPLC at room temperature. In short, rotation about the N-C7' axis of 4 is similar to that of 1. To our surprise, however, 4 could be resolved by chiral HPLC (CHIRALPAK IB) into enantiomers at room temperature, indicating that it exists as a racemate of the atropisomers. It is clear that the steric hindrance provided by the C-2' and C-6' substituents forms a higher rotational barrier for the C7'-C1' axis than for the N-C7' axis. It should be noted that each cis-like and trans-like conformer of 4 exists as the racemate of a R and a S isomers, as shown in Figure 3.

We managed to obtain each enantiomer of 4 using preparative chiral HPLC at about 96% ee, with high stereochemical stability at room temperature (see Supporting Information). It was confirmed that each enantiomer exists as an equilibrium mixture of cis-like and trans-like conformers. The isolated atropisomers have opposite $[\alpha]_D$ values: 4A (with shorter retention time in HPLC using CHIRALPAK IB) as 96% ee showed $\left[\alpha\right]^{24}$ D = 25.7 (c 0.1, CHCl₃) and **4B** (with longer retention time in HPLC using CHIRALPAK IB) as 96% ee showed $\left[\alpha\right]^{23}$ +23.1 (c 0.15, $CHCl_3$), confirming that they are enantiomers. The isolation of each enantiomer with high stereochemical stability provides evidence that rotation about the C7'-C1' axis can be fully restricted by substituents at C-2' and C-6'. It is worth noting that the N-C7' axis can allow rotation even though the neighboring C7'-C1' axis is blocked to



Figure 3. Conformers including atropisomers of the indometacin analogue 4.

rotation. Judging from the above, these two axes behave independently for **4** in solution. To the best of our knowledge, this type of atropisomer has not been previously reported. The separated enantiomers of **4** (**4A** and **4B**) were hydrolyzed to afford the corresponding carboxylic acid derivatives **5** (**5A**, **5B**) (Scheme 2). We fortunately succeeded in obtaining the enantiomer **5A** as a single crystal.

Scheme 2. Separation of 4 into Atropisomers 4A and 4B and Hydrolysis to the Corresponding Carboxylic Acids 5A and 5B



On the basis of its X-ray analysis, **5A** was assigned to be (a*S*), and hence, **5B** to be (a*R*) (see Supporting Information). We examined the stereochemical stability of the enantiomers (**5A** and **5B**) and found that it was estimated to be high: racemization occurred after storage for 48 h at 37 °C in EtOH. Thus, the activation free-energy barrier to rotation (ΔG^{\dagger}) was calculated to be 106 kJ/mol²⁸ (see Supporting Information).

We next examined the activities of the racemate **5** and each pure enantiomer (**5A** and **5B**) against COX-1/2

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Table 1. In	Vitro Cyclooxygenase	Assay	of 5 (Racemate and
Atropisome	ers)		

		$IC_{50}\left(\mu M\right)$		
	$\left[\alpha\right]^{22}{}_{D}\left(CHCl_{3}\right)$	COX-1	COX-2	
5 5A (aS)	-21.2(c 0.13, as 96% ee)	13.4 ± 3.4 >200	>500 >500	
5B (a <i>R</i>)	$+20.4 (c \ 0.10, as \ 94\% \ ee)$	15.6 ± 1.8	>500	
indometacin		0.106 ± 0.042	3.50 ± 041	

(Table 1). It was found that the racemate 5 or enantiomers 5A and 5B did not show COX-2-inhibitory activity at a concentration of 500 μ M. On the other hand, the racemate 5 exhibited potent COX-1-selective inhibitory activity with an IC₅₀ value of 13.4 \pm 3.4 μ M. The result is attractive because COX-1 inhibitors have been revaluated recently.¹⁴ It appears reasonable to assume that the COX-1 versus COX-2 selectivity of 5 is closely linked with its conformation. As pointed out above, there is a difference in the conformation of indometacin relevant to the interactions with COX-1 and COX-2. Although the interaction of indometacin with COX-2 has been determined to be only the cis conformation,¹³ indometacin interacts with COX-1 in the cis or trans conformation.¹² On the basis of ¹H NMR spectroscopy, compound 5 exists as an equilibrium mixture of cis-like and trans-like conformation (cis-like:translike = 1:1.6) in CDCl₃. Such an equilibrium biased toward the trans-like conformer should be advantageous to COX-1.

In addition, it was revealed that the enantiomer **5B** (a*R*) shows potent activity against COX-1 with an IC₅₀ value of $15.6 \pm 1.8 \,\mu$ M. The other enantiomer **5A** (a*S*) showed no inhibitory activity at a concentration of 200 μ M. The results clearly indicate that COX-1 recognizes the stereochemistry of the axial chirality arising from the C7'-C1' axis. We assumed that C-2' and C-6' disubstituted indole derivative **5** provided an atropisomeric property that should be a clue for the design of COX-1/2-selective indometacin derivatives.

In conclusion, one of the active conformations of indometacin for COX-1 among the numerous possible conformations has been revealed by the introduction of different substituents at the 2'- and 6'-positions of indometacin (twisted conformation). This is the first example of introducing atropisomerism to N-benzoylated indole derivatives. We hope that our newly discovered atropo-selective bioactivity triggers the development of pure atropisomers as new drug candidates in medicinal chemistry.

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Supporting Information Available. NMR data of 1–5, NOESY data of 2, analytical study of enantiomers of 4 (4A and 4B) and 5 (5A and 5B), stereochemical stability of the enantiomers of 5 (5A and 5B), crystal data (CIF file) for 3 and 5A, in vitro cyclooxygenase assay. This material is available free of charge via the Internet at http://pubs. acs.org.